

Available Online at eSci Journals

# eSci Journal of Plant Pathology

ISSN: 2305-106X (Online), 2306-1650 (Print) http://www.escijournals.net/EJPP



# IMPROVMENT BIOCONTROL OF DAMPING-OFF AND ROOT ROT/WILT OF FABA BEAN BY SALICYLIC ACID AND HYDROGEN PEROXIDE

Montaser F. Abdel-Monaim\*

Plant Pathology Research Institute, Agriculture Research Center, Giza 12619, Egypt.

#### ABSTRACT

Rhizoctonia solani, Fusarium solani, F. oxysporum and Macrophomina phaseolina were found to be associated with root rot and wilt symptoms of faba bean plants collected from different fields in New Valley governorate, Egypt. All the obtained isolates were able to attack faba bean plants (cv. Giza 40) causing damping-off and root rot/wilt diseases. R. solani isolates 2 and 5, F. solani isolate 8, F. oxysporum isolate 12 and M. phaseolina isolate 14 were the more virulent ones in the pathogenicity tests. Biocontrol agents (Trichoderma viride, Bacillus megaterium) and chemical inducers (salicylic acid and hydrogen peroxide) individually or in combination were examined for biological control of damping-off and root rot/wilt and growth promotion of faba bean plants in vitro and in vivo. Both antagonistic biocontrol agents and chemical inducers either individually or in combination inhibited growth of the tested pathogenic fungi.Biocontrol agents combined with chemical inducers recorded the highest inhibited growth especially in case SA + T. viride and SA+ B. megaterium. Under green house and field conditions, all treatments significantly reduced damping-off and root rot/ wilt severity and increased survival of plants. Also, these treatments increased fresh and dry weights of the survived plants in pots compared with control. The combination of biocontrol agents and chemical inducers were more effective than using them individually and SA+ T. viride was the best treatment in this respect. Under field conditions, all these treatments significantly increased growth parameters (plant height and No. of branches plant-1) and yield components (No. of pods and seedsplant-1, weight of 100 seeds and total yield feddan-1 and protein content in both seasons (2010-2011 and 2011-2012). Faba bean seeds soaked in SA+ T. viride and SA+ B. megaterium were recorded the highest growth parameters and yield components. Generally, the combination of biocontrol agents and chemical inducers recoded the best results for controlling damping-off and root rot/wilt diseases in greenhouse and field with addition improved plant growth and increased yield components in the field.

**Keywords**: Faba bean, Root rot and wilt, Salicylic acid, Hydrogen peroxide, Biological control, Growth and yield parameters.

# **INTRODUCTION**

Faba bean (*Vicia fabe* L.) is used as an important human food in developing countries and as an animal feed, mainly for pigs, horses, poultry and pigeons in industrialized countries. Feeding value of faba bean is high and this legume has been considered as a meat extender or substitute due to its high protein content (20-41%) (Chaven *et al.*, 1989). Root rot and wilt diseases caused by several soil borne fungal pathogens are widespread and serious in many crops cultivated in different soil types. Faba bean is subjected to attack by many pathogenic organisms wherever the crop is grown. Several root rot and wilt pathogens such as

\* Corresponding Author:

Email: fowzy\_2008@yahoo.com

© 2012 eSci Journals Publishing. All rights reserved.

Rhizoctonia solani, Fusarium solani, and Macrophomina phaseolina are reported to attack faba bean roots and stem base causing serious losses in seed germination and plant stand (Abdel-Kader *et al.*, 2011).

An investigation for controlling such diseases is considered important, especially in view of their wide prevalence in Egypt. Several attempts to control root rot and wilt diseases could be accepted. However, fungicides are considered one of several factors involving in environmental pollution, in spite of their satisfactory results in the control of plant diseases. In addition, control of disease with fungicides has proven very difficult, and almost all fungicides are effective only at phytotoxic levels (Jarvis, 1988). Recently, the growing concern over the use of pesticides to human health and environment has brought increasing interest in the use

of alternatives characterized with negative impact on the environment. Therefore, present activity focuses on finding compounds that are safe to human and environment. An alternative to fungicidal application, it may be possible to utilize a scheme of inducible plant defenses which provides protection against a broad spectrum of disease-causing organisms. Among synthetic inducers, salicylic acid (SA) and hydrogen peroxide (H2O2) have been found to be active as antimicrobial agents in various trials as disease resistance inducers. These have been reported for inducing resistance against several plant pathogens, i.e. F. oxysporum in tomato (Abdel- Monaim et al., 2012), R. solani in faba bean and potato (Metwally, 2004), F. solani in lentil (Morsy, 2005) and M. phaseolina in water melon(Saleh et al., 2007). On the other hand, the application of biological control using antagonistic microorganisms proved to be successful for controlling various plant diseases in many countries. Biological control is proposed to be an effective and nonhazardous strategy to reduce crop damage caused by plant pathogens. In recent years the B. megaterium and T. viride have been extensively used for plant growth promotion and disease control (Abdel-Monaim, 2010). Biological control of soil borne pathogens is often attributed to improved nutrition that boosts host defenses or to direct inhibition of pathogen growth and activity. Amendment with certain abiotic factors (inducers) appears to stimulate the disease resistance by indirectly stimulating indigenous populations of microorganism that are beneficial to plant growth and antagonistic to pathogens. For example chitin amendment of soil has been found to stimulate the growth of chitinolytic microorganisms (De Boer et al., 1999), increase the biocontrol activity and stimulate the expression of plant defense proteins (Roby et al., 1987). All these effects may culminate in enhancing plant protection. Similarly, salicylic acid (SA) and H<sub>2</sub>O<sub>2</sub> amendment was tested in combination with biocontrol agents. Saikia et al. (2003) tested the efficiency of P. fluorescence with or without SA amendment in chickpea against Fusarium wilt infection. The application of P. fluorescence (pf4-92) with SA recorded highest protection of chickpea seedlings against wilting.

Thus, the aim of this study is to (i) screen biocontrol agents and chemical inducers individually or in combination capable of inhibiting the growth of *R. solani, F. solani, F. oxysporum* and *M. phaseolina*, (ii) determine the biocontrol efficiency of *B. megaterium* 

and T. viride, alone and in combination with chemical inducers ( $H_2O_2$  and SA) amendment against damping-off and root rot/wilt diseases of fababean and (iii) determine the effect of B. megaterium and T. viride, alone and in combination with chemical inducers ( $H_2O_2$  and SA) amendment on promoting growth and yield components of fababean in field.

## **MATERIALS AND METHODS**

Source of Faba Bean Seeds and Growth of Plants: Faba bean (*V. fabae* L.) cultivar Giza 40 used in this study, was obtained from Legume Crop Research Department, Field Crop Res. Inst., Agric. Res. Center, Ministry of Agriculture, Egypt. Seeds were planted in plastic pots with 30 cm diameter (2.4 kg soil), filled with a pasteurized mixture of soil and sand (4:1 w/w). Six seeds were sown in each pot and these pots were irrigated every three days.

Isolation, Purification and Identification of the Causal Organism (s): Samples of faba bean plants showing root rot and wilt symptoms were collected from different farms located in New Valley governorate. The infected roots were thoroughly washed with running tap water, cut into small fragments, superficially sterilized with sodium hypochlorite (5%) for 2 minutes, washed several times with sterile distilled water and dried between sterilized filter paper. The sterilized pieces were transferred into potato dextrose agar (PDA) medium supplemented with penicillin (20 Iu ml-1) and incubated at 25 ±1 °C, then examined daily for fungal growth. The fungal colonies were purified using single spore or hyphal tip techniques suggested by Booth (1985) and Dhingra and Sinclair (1985) and then identified according to their morphological and microscopical characters described by Booth (1985) and Barnett and Hunter (1986).Identification was confirmed by Assiut Mycological Centre (AUMC), University University, Assiut, Egypt. The obtained isolates were maintained on PDA slants and kept in refrigerator at 5°C for further study.

**Pathogenicity Tests:** The pathogenicity tests of the obtained fungal isolates carried out at New Valley Agric. Res. Station, on Giza 40 faba bean cultivar in pots containing soil infested with the obtained isolates individually using homogenized culture technique according to Muthomi *et al.* (2007).

**Preparation of the Fungal Inocula:** The inocula of the obtained isolates were prepared from one week old culture grown on 50 ml potato dextrose broth medium

(PDB) in conical flask (250 ml) and incubated at 25±1 °C. The content of flask were homogenized in a blender for one min. Plastic pots were filled with sterilized soil and mixed with fungal inocula at rate 100 ml homogenized culture per pot, seven days before planting. Five pots were used as replicates per isolate and another 5 pots with equal amount of sterile PDB medium without fungal inoculation were applied as control. Five sterilized surface seeds were sown in each pot. Percentage of damping-off was recorded 30 days after planting, respectively. While severity of root rot / wilt was determined after 90 days according to Abdou et al. (2001) using a rating scale of 0 to 5 on the basis of root discoloration or leaf yellowing as follows, 0 = neither root discoloration nor leaf yellowing, 1= 1-25% root discoloration or one leaf yellowed, 2= 26-50% root discoloration or more than one leaf yellowed, 3= 51-75% root discoloration plus one leaf wilted, 4= up to 76% root discoloration or more than one leaf wilted, and 5= completely dead plants. For each replicate a disease severity index (DSI) similar to that one described by Liu et al. (1995) was calculated as follows:

$$DSI = \frac{\Sigma d}{dma \times n} \times 100$$

Whereas: d is the disease rating possible, d max is the maximum disease rating and n is the total number of plants examined in each replicate.

In vitro Studies: The used antagonistic organisms were Trichoderma viride isolate TVM2 and Bacillus megaterium isolate BSM5. These isolates provided by Dr. Montaser, F. Abdel-Monaim Plant Pathol. Res. Inst., Agric. Res. Center (Abdel-Monaim, 2010). While, chemical inducers used were salicylic acid (SA) and hydrogen peroxide ( $H_2O_2$ ) at concentration 4 mM and 1 mM, respectively. The effect of biocontrol agents and chemical inducers individually and/or in combination on faba bean pathogenic fungi was studies as follows:

A) Antagonistic Biocontrol Agents: *Trichoderma viride* and the tested pathogenic fungi (*R. solani, F. solani* and *M. phaseolina*) were cultured on PDA medium for 7 days at 25±1 °C. A disc (0.7 cm diameter) of the antagonistic fungal colony was cut and placed opposite to the colony of the pathogenic fungal isolates on PDA medium. On the other hand, *Bacillus megaterium* isolate was streaked at opposite ends of PDA plates near edge and incubated at 25±1 °C for 24 hr. Then a mycelial disc (0.7 cm) of the tested fungi was placed in the center of each plate.

- B) Chemical Inducers: The effect of chemical inducers on the growth of pathogenic fungi was evaluated in PDA medium. 20 ml of PDA medium containing 4 mM SA, 1 mM  $H_2O_2$  was poured in the plates and inoculated with the pathogenic fungi.
- C) Combination Between of Biocontrol Agents and Chemical Inducers: Flasks (250 ml) containing 200 ml PDA medium were amended with 4 mM SA and 1 mM  $\rm H_2O_2$  individually, then each flask was poured in 10 plates. These plates were inoculated with antagonistic isolates and pathogenic isolates as before. For control treatment, the agar plug of only pathogen isolates was placed on PDA plates. The inoculated plates incubated at 25±1 °C until colony of control grew to full plate. At this point, colony diameter was measured using ruler. Percentage of growth inhibition of pathogen was calculated using the formula below:

% Inhibition =  $(A-B)/A \times 100$ 

Where:

A = Colony diameter of pathogen in control B = Colony diameter in treated plates

#### In vivo Studies

**Greenhouse Experiments:** The effects of biocontrol agents (T. viride,B. megaterium) and chemical inducers (SA,  $H_2O_2$ ) individually or in combination against faba bean damping-off and root rot diseases incited by R. solani, F. solani and M. phaseolina were evaluated under greenhouse conditions. Trichoderma viride grown on PDA at  $25\pm1$  °C for 7 days was prepared in sterilized water to  $10^6$ cfu /ml. Cell suspension of B. megaterium grown on nutrient broth medium for 3 days at  $25\pm1$  °C were adjusted to  $2.5\times10^8$ cfu /ml. SA and  $H_2O_2$  were prepared as solutions (water or biotic suspension) at 4 and 1 mM, respectively. The combination between biocontrol agents and chemical inducers prepared with dissolving chemical inducers in suspension of biocontrol agents.

Faba bean seeds soaked for 6 hr. in the following treatments.1- SA (4 mM), 2-  $H_2O_2$  (1 mM), 3- T. viride ( $10^6$ cfu/ml), 4 – B. megaterium ( $2.5\times10^8$ cfu /ml), 5- SA+ T. viride, 6- SA+ B. megaterium, 7-  $H_2O_2$ + T. viride, 8-  $H_2O_2$ + B. megaterium, 9- control. Plastic pots were filled with sterilized soil and mixed with fungal inocula at rate 100 ml homogenized culture per pot, seven days before planting and then were sown by 6 seeds of each treatment. Five replicates were used for each treatment. In control treatment, faba bean seeds soaked in water for 6 hrs. and sown at same rate. Also, treated and untreated seeds were sown in pots containing

uninfested soil to study the effect of these treatments on plant growth. Pots were irrigated as needed.

All pots were examined after 30 and 90 days to record the percentage of damping-off and root rot/wilt severity, respectively. Also, the end of experiment, the survived plants were weighted to record fresh weight per plant then dried at 80  $^{\circ}$ C for 24 hr to record the dry weight per plant.

Field Experiments: Field experiment was carried out at New Valley Res. Station Farm during 2009-2010 and 2010-2011 growing seasons, to evaluate the efficiency of the tested biocontrol agents (*T. viride, B. megaterium*) and chemical inducers (SA and H<sub>2</sub>O<sub>2</sub>) individually or in combination for controlling damping-off and root rot diseases and its effect on growth and yield parameters under field conditions. The experimental design was a complete randomized block with three replicates. The experimental unit area was 10.5 m<sup>2</sup> (3.5 × 3 m). Each unit included 5 rows; each row was 3.5 m in length and 60 cm width. Faba bean seeds (cv. Giza 40) were soaked in treatments described above for 6 hr The seeds treated were sown in hills 25 cm apart on both sides of 6 cm ridges in both seasons, 2 seed per hill. In control treatment, faba bean seeds were soaked in water for 6 hr and were sown in the same rate. The normal cultural practices of growing faba bean were followed. Percentages of damping-off and root rot/wilt severity were recorded 30 and 90 days after sowing. At harvest, plant height (cm), number of branches, pods and seeds plant<sup>-1</sup>, 100- seed weight and total yield (kg feddan <sup>-1</sup>) were measured. Protein percentage content in seeds was recorded using the method of Jackson (1973).

**Statistical Analysis:** All experiments were performed twice. Analysis of variance was carried out using MSTAT-C, 1991 program version 2.10. Least significant difference (LSD) was employed to test for significant difference between treatments at  $P \le 0.05$  (Gomez and Gomez, 1984).

## **RESULTS**

## **Isolation and Identification of the Causal Organisms:**

Twenty isolates of different soil-borne fungi listed in Table (1), were isolated from wilted and rotten roots of faba bean plants, showed root-rot and wilt symptoms, cultivated in different fields in New Valley governorate. These isolates were identified as *Rhizoctonia solani* (5 isolates), *Fusarium solani* (4 isolates), *F. oxysporum* (4 isolates), *Macrophomina phaseolina* (4 isolates) as well as 3 isolates not identified.

**Pathogenicity Tests:** The obtained isolates were tested on faba bean cv. 40 under greenhouse conditions in pots. Results presented in Table (1) reveal that all tested isolates could infect the roots of faba bean and cause damping-off, root rot and wilt symptoms thus reduce the survived plants. The highest damping-off infection (60%) was recorded by *R. solani* (isolate 2) followed by *F. solani* isolate 8 (56%) then *R. solani* isolate 3 and *F. solani* isolate 9, where recoded 52% damping –off. On the other hand, all tested isolates caused root rot/wilt ranging between 2.59- 50.67%. *Fusarium oxysporum* isolates 12 and 13 followed by *R. solani* isolate 5 caused the highest root rot/wilt severity, whereas recorded 50.67, 44.33 and 39.67%, respectively.

It is concluded from results that *R. solani* isolates 2 and 5, *F. solani* isolate 8, *F. oxysporum* isolate 12 and *M. phaseolina* isolate 14 were the more virulent ones whereas recorded the lowest survived plants (4.33, 16.33, 10.4, 13.33 and 21.33%, respectively).

As mentioned before in the pathogenicity test experiments, the highly pathogenic fungal isolates obtained from faba bean roots *i.e.* Rhizoctonia solani (2), Fusarium solani (8), F. oxysporum (12) and M. phaseolina (14) were chosen to complete the further studies based on their pathogenic abilities.

# Effect of Biocontrol Agents and Chemical Inducers on Damping-off and Root Rot/Wilt, Fresh and Dry Weights under Greenhouse Conditions

**A) Diseases Incidence:** Data presented in Table 3 show that all treatments significantly reduced damping-off and root rot/wilt severity and increased survived plants of faba bean which have been artificially infested with tested pathogens *R. solani, F. solani, F. oxysporum* and *M. phaseolina*, compared to untreated control treat

ment. Soaking seeds in biocontrol agents combined with chemical inducers reduced damping –off and root rot/wilt severity more than used any of them individually. Chemical inducers were more effective to reduce damping-off and root rot/wilt than biocontrol

agents. On the other hand, the combination of SA and *T. viride* recorded the highest protection against to any of the tested fungi followed by SA+ *B. megaterium* while seed soaked in *B. megaterium* or *T. viride* recorded the lowest ones.

Table 1. Pathogenicity of fungi isolated from root of faba bean diseased plants.

Isolates No.	% Damping-off	% Root rot/wilt	% Survived plants
	Rhizoc	tonia solani	
1	18 j	25.33 f	56.67 e
2	60 a	35.67 d	4.33 n
3	52 c	14.33 h	33.67 i
4	44 d	12.69 hi	43.31 g
5	44 d	39.67 c	16.33 l
	Fusar	ium solani	
6	24 i	12.97 hi	63.03 d
7	28 h	20.33 g	51.67 f
8	56 b	33.6 d	10.4 m
9	52 c	20.14 g	27.86 j
	F. ox	ysporum	
10	32 g	10.00 i	58 e
11	40 e	35.00 d	25 j
12	36 f	50.67 a	13.3 lm
13	28 h	44.33 b	27.67 j
	Macrophor	nina phaseolina	
14	50 c	28.67 e	21.33 k
15	28 h	12.67 hi	59.33 e
16	40 e	22.54 fg	37.46 h
17	20 j	12.56 hi	67.44 c
	Oth	er fungi	
18	81	2.59 j	89.41 a
19	4 m	5.64 j	90.36 a
20	12 k	5.28 j	82.72 b

Different letters indicate significant differences among treatments within the same column according to least significant difference test ( $P \le 0.05$ ).

Table 2. Effect of biocontrol agents and chemical inducers individually or combined on growth of pathogenic tested fungi in vitro.

Treatments	Inhibition of linear growth (%)								
Treatments	R. solani	F. solani	F. oxysporum	M. phaseolina					
Trichoderma viride	38.47 d	55.58 c	44.69 d	60.14 b					
Bacillus megaterium	32.33 e	49.22 d	40.23 e	46.20 d					
Salicylic acid (SA)	8.87 f	13.56 e	$10.55\mathrm{g}$	11.58 f					
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	11.36 f	16.25 e	14.00 f	15.58 e					
SA+ T. viride	57.00 a	70.54 a	62.35 a	64.30 a					
SA+ B. megaterium	46.41 b	63.25 b	58.65 b	56.36 c					
$H_2O_2+$ T. viride	42.24 c	59.25 bc	52.31 c	56.14 c					
H <sub>2</sub> O <sub>2</sub> + B. megaterium	40.14 cd	57.47 c	44.96 d	53.34 c					

Different letters indicate significant differences among treatments within the same column according to least significant difference test ( $P \le 0.05$ ).

Table 3. Effect of seed soaking in biocontrol agents and chemical inducers individually or combination for controlling damping-off, root rot and survival plants under greenhouse conditions.

	R. solani			F. solani			F. oxysporum			M. phaseolina		
Treatments	% Damping- off	% Root rot	% Survival plants	% Damping -off	% Root rot	% Survival plants	% Damping -off	% Root rot	% Survival plants	% Damping -off	% Root rot	% Survival plants
Trichoderma viride	24 c	6.59 de	69.41 c	24 b	8.67 bc	67.33 e	2 b0	18.23 b	61.77 e	12 c	8.67 bc	79.33 cd
Bacillus megaterium	28 b	11.7 b	60.3 d	20 c	10.23 b	69.77 e	2 b0	20.23 b	59.77 e	1 b6	10.33 b	73.67 d
Salicylic acid (SA)	16 g	7.67 g	76.33 f	12 h	6.47 f	81.53 g	12 f	12.36 f	75.64 g	8 f	6.67 e	85.33 f
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	20 d	8.9 c	71.1 c	16 d	7.25 cd	76.75 d	16 c	15.56 с	68.44 d	8 d	10.33 b	81.67 bc
SA+ T. viride	8 f	4.33 f	87.67 a	4 g	3.6 e	92.4 a	4 f	6.36 e	89.64 a	e4	5.6 d	90.4 a
SA+ B. megaterium	12 e	5 ef	83 ab	8 f	5.67 d	86.33 b	8 e	8.23 de	83.77 b	6 de	6.33 d	87.67 ab
$H_2O_2+$ T. viride	12 e	6.58 de	81.42 b	12 e	6.00 d	82.00 c	12 d	9.36 d	78.64 c	6 de	6.33 d	87.67 ab
$H_2O_2$ + B. megaterium	12 e	6.89 d	81.11 b	8 f	6.00 d	86.00 b	8 e	10.25 d	81.75 bc	6 de	8.33 c	85.67 abc
Control	60 a	26.54 a	13.46 e	52 a	32.33 a	15.67 f	32 a	50.25 a	17.75 f	48 a	33.52 a	18.48 e

Different letters indicate significant differences among treatments within the same column according to least significant difference test ( $P \le 0.05$ ).

Table 4. Effect of seed soaking in biocontrol agents and chemical inducers individually and/or combination on fresh and dry weight (gm plant-1) of root and shoot healthy and infected faba bean plants.

	Uninfe	ected		Infected with;							
Treatments	ENA	DM	R. solani		F. solani		F. oxysporum		M. phaseolina		
	FW	DW	FW	DW	FW	DW	FW	DW	FW	DW	
Trichoderma viride	20.66 cd	4.38 c	17.59 bc	3.58 bc	16.85 cde	3.96 bcd	14.36 cd	3.37 bc	17.72 b	3.74 b	
Bacillus megaterium	20.01 cd	4.22 c	15.99 cd	3.24 cd	15.21 de	3.47 de	13.89 de	3.21 c	16.56 b	3.56 b	
Salicylic acid (SA)	18.66 f	3.94 f	14.84 f	3.01 f	12.28 g	2.69 g	10.55 g	2.36 e	10.50 d	2.22 d	
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	17.30 de	3.55 d	13.63 d	2.71 d	14.21 e	3.16 e	11.66 e	3.11 c	10.07 с	2.04 c	
SA+ <i>T. viride</i>	26.90 a	5.43 a	21.2 a	4.14 a	21.12 a	4.72 a	18.36 a	3.96 a	22.28 a	4.81 a	
SA+ B. megaterium	24.62 ab	5.24 a	20.17 ab	3.84 ab	20.02 ab	4.45 ab	17.45 ab	3.85 ab	19.27 b	4.19 ab	
$H_2O_2+$ T. viride	22.54 bc	4.73 b	18.43 abc	3.53 bc	19.03 abc	4.15 bc	16.56 abc	3.58 abc	18.42 b	4.01 b	
$H_2O_2$ + B. megaterium	21.91 bc	4.53 bc	16.85 с	3.21 cd	17.53 bcd	3.82 cd	15.42 bcd	3.37 bc	18.15 b	3.76 b	
Control	15.27 e	3.05 e	9.32 e	1.77 e	8.08 f	1.72 f	6.45 f	1.38 d	9.08 c	1.84 c	

Different letters indicate significant differences among treatments within the same column according to least significant difference test ( $P \le 0.05$ ).

FW, Fresh weight; FD, Dry weight

Table 5. Effect of seed soaking in biocontrol agents and chemical inducers individually and/or combination on controlling damping-off, root rot/wilt and survival plants under field conditions during growing seasons 2010-2011 and 2011-2012.

	S	eason 2010	-2011	Season 2011-2012			
Treatments	% Damping- off	Damping- % %  Root rot Survival plan		% Damping-off	% Root rot	% Survival plants	
Trichoderma viride	13.33 с	10.33 b	76.34 ef	12.00 с	9.50 b	78.50 ab	
Bacillus megaterium	18.00 b	10.00 bc	72.00 f	15.67 b	9.33 b	75.00 bc	
Salicylic acid (SA)	9.67 d	6.67 de	83.66 cd	8.00 de	7.90 bc	84.10 ab	
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	10.67 cd	7.96 cd	81.37 de	10.00 cd	6.33 cd	83.67 ab	
SA+ T. viride	4.00 e	4.00 f	92.00 a	3.67 g	3.00 e	93.33 a	
SA+ B. megaterium	5.67 e	5.26 ef	89.07 ab	5.00 fg	4.84 d	90.16 ab	
$H_2O_2$ + T. viride	7.33 de	4.33 f	88.34 abc	6.33 ef	5.00 d	88.67 ab	
H <sub>2</sub> O <sub>2</sub> + <i>B. megaterium</i>	9.67 d	5.50 ef	84.83 bcd	8.00 de	5.30 d	86.70 ab	
Control	32.33 a	15.40 a	52.27 g	28.00 a	13.00 a	59.00 c	

Different letters indicate significant differences among treatments within the same column according to least significant difference test ( $P \le 0.05$ ).

B) Fresh and Dry Weight: In the present study, biocontrol agents (T. viride and B. megaterium) and chemical inducers (SA and  $H_2O_2$ ) individually or combined recorded significantly increased fresh and dry weights either in presence or absence ofpathogenic fungi (Table 4). Biocontrol agents combined with chemical inducers increased fresh and dry weights more than using any of them individually. Also, biocontrol agents gave fresh and dry weights higher than usage of chemical inducers. Generally, combination between SA and T. viride recorded the highest fresh and dry weights of survived plants either in uninfested or infested soils with pathogenic fungi followed by SA+B. megaterium. On other hand,  $H_2O_2$  recorded the lowest fresh and dry weights in this respect.

Effect of Biocontrol Agents and Chemical Inducers on Disease Incidence under Field Conditions: The efficacy of chemical inducers and biocontrol agents individually and/or in combination against the incidence of damping-off and root rot/wilt diseases of faba bean, were evaluated under field conditions as seed soaking. Data in Table 5 clearly demonstrate that all treatments significantly reduced damping-off and root rot/wilt severity compared with the control. Biocontrol agents with chemical inducers were more effective than being used alone. Also, the obtained data show that seed soaking in any of the chemical inducers (SA or H<sub>2</sub>O<sub>2</sub>) was more effective to reduce damping-off and root rot/wilt than any of the biocontrol agents. SA+T. viride recorded the highest reduction of damping-off and root rot/wilt in both seasons, whereas recorded

92% and 93.33% survived plants compared with 52.27 and 59% in control plants in both seasons, respectively. In the contrary, faba bean seeds soaked in *B. megaterium* or *T. viride* were recorded the lowest ones in both seasons (72, 75% survival plants in first seasons and 76.34, 78.50% in second season, respectively).

Effect of Biocontrol Agents and Chemical Inducers on Growth Parameters, Yield Components and Protein Content under Field Conditions: The effect of faba bean seed soaked in biocontrol agents (T. viride, B. megaterium), chemical inducers (SA, H2O2) on growth parameters and yield components under field conditions during seasons 2010-2011 and 2011-2012 were studied. Data in Table 6 indicate that seed the combination between and biocontrol agents and chemical inducers significantly improvement faba bean growth and branching more than used individually. Also, seed soaked in any of biological agents were more effective in this respect than used any chemical inducers. The combination between SA and T. viride were the most effective treatments; they recorded the highest plant height (122.6, 132.4 cm) and number of branches (4.1 and 4.2 branch plant<sup>-1</sup>) in both seasons, respectively. Also, this treatment recorded the highest yield components i.e. number of pods (19.3, 20.9 pods plant<sup>-1</sup>), number of seeds (47.3, 53.2 seed plant<sup>-1</sup>), weight of 100 seed (83.9, 85.8 gm) and total seed yield (1851, 1986 kg feddan-1) in both seasons, respectively. On the contrary, faba bean seed soaked in H<sub>2</sub>O<sub>2</sub> was less effective in both seasons compared the other treatments. On the other hand, the obtained data show

that all treatments significantly increased protein content in seeds compared with untreated control. Also, SA +T. viride was recorded the highest protein content (30.8 and 31%) followed by SA + B. megaterium (29.7, 30.1%) in both seasons, respectively. While,  $H_2O_2$  individually was recorded the lowest increased in this respect.

#### **DISCUSSION**

Faba bean (*Vicia faba* L.) is one of the most important legume crops. It is infected with many fungal pathogens causing considerable yield losses where damping-off, root-rot, wilt diseases are the most important fungal diseases affecting faba bean production in Egypt (Abdel-Kader *et al.*, 2011).

Sum of 20 isolates of different soil-borne fungi were isolated from roots of faba bean plants showed root-rot and wilt symptoms collected from different fields growing in New Valley governorate. These isolates were identified as *Rhizoctonia solani, Fusarium solani, F. oxysporum and Macrophomina phaseolina*. Pathogenicity test of the obtained isolates revealed that all tested isolates could infect the roots of faba bean (cv. Giza 40) causing damping-off, root rot/wilt and reduced survived plants. *R. solani* isolates 2 and 5, *F. solani* isolate 8, *F. oxysporum* isolate 12 and *M. phaseolina* isolate 14 were the more virulent ones. These results are similar to those obtained by Metwally (2004) and Abdel-Kader *et al.*, (2011).

Table 6. Effect of seed soaking in salicylic acid, hydrogen peroxide, Trichoderma viride, Bacillus subtilis individually and/or combination on growth and yield parameters under field condition during growing seasons 2010-2011 and 2011-2012.

	Season 2010-2011									
Treatments	Plant height (cm)	No. of branches plant <sup>-1</sup>	No. of pods plant-1	No. of seed plant <sup>-1</sup>	Weight of 100 seeds	Total yield fed <sup>1</sup>	% Protein			
Trichoderma viride	110.3 cd	3.2 cd	16.4 bc	41.5 cd	80.9 a	1551.4 bcd	28.3 abc			
Bacillus megaterium	104.0 de	3.0 d	14.5 d	37.5 ef	81.2 a	1453.0 de	27.9 bc			
Salicylic acid (SA)	98.5 e	3.3 cd	15.0 cd	36.4 f	81.6 a	1531.5 cd	27.7 bc			
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	89.2 f	2.9 d	13.6 de	34.8 f	79.9 a	1393.3 e	26.8 c			
SA+ T. viride	126.0 a	4.1 a	19.3 a	47.3 a	83.9 a	1851.0 a	30.8 a			
SA+ B. megaterium	119.6 b	3.9 ab	17.4 b	43.2 bc	82.5 a	1633.0 bc	29.7 ab			
$H_2O_2+$ <i>T. viride</i>	115.3 bc	3.9 ab	17.3 b	44.9 ab	82.9 a	1662.5 b	28.1 abc			
H <sub>2</sub> O <sub>2</sub> + B. megaterium	109.5 cd	3.6 bc	16.5 b	39.2 de	82.1 a	1547.0 bcd	28.0 bc			
Control	70.3 g	2.3 e	12.5 e	26.9 g	72.4 b	1060.0 f	26.1 c			
		Season	2011-201	2						
Trichoderma viride	117.2 cd	3.2 c	17.0 bc	43.2 cd	82.1 a	1644.5 cd	28.6 bc			
Bacillus megaterium	111.5 de	3.2 c	15.3 cd	38.7 ef	83.6 a	1537.2 de	27.4 cd			
Salicylic acid (SA)	103.5 f	3.4 c	17.3 bc	41.3 de	84.4 a	1604.0 d	28.3 bc			
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	89.3 g	2.8 d	15.8 c	36.5 f	82.6 a	1475.8 e	27.4 cd			
SA+ T. viride	132.4 a	4.2 a	20.9 a	53.2 a	85.8 a	1986.0 a	31.0 a			
SA+ B. megaterium	126.3 b	3.9 ab	18.8 ab	47.3 b	84.8 a	1758.7 b	30.1 ab			
H <sub>2</sub> O <sub>2</sub> + <i>T. viride</i>	118.5 с	4.1 ab	18.6 b	46.2 bc	84.2 a	1729.0 bc	28.7 abc			
H <sub>2</sub> O <sub>2</sub> + B. megaterium	110.4 e	3.8 b	17.3 bc	39.9 def	84.8 a	1552.5 de	27.8 bc			
Control	73.5 h	2.4 e	13.5 d	32.1 g	75.6 b	1202.8 f	25.2 d			

Different letters indicate significant differences among treatments within the same column according to least significant difference test ( $P \le 0.05$ ).

Sustainable farming systems strive to minimize the use of synthetic pesticides and to optimize the use of alternative management strategies to control soil-borne pathogens (Harrier and Watson, 2004). Antagonistic fungi and bacteria in plant root zone are a key agent of change in soil agroecosystems. Interactions between

plant root systems and rhizobacteria have a profound effect on crop health, yield and soil quality. Root zone antagonistic fungi and bacteria are able to generate a wide array of secondary metabolites which can have a positive influence on plant growth; enhancing the availability of minerals nutrients, improving nitrogen

fixation ability, decreasing susceptibility to frost damage, improving plant health through the biocontrol of phytopathogens, inducing systemic plant disease resistance and facilitating plant establishment, growth and development (Sturz and Christie, 2003). Also, acquired resistance by using abiotic-agents as inducers seems to be one of alternatives to substitute for, or at least to decrease the use of fungicides in plant disease control. Excessive and improper use of pesticides including fungicides presents a menace to the health of human, animal and environment (Guzzo et al., 1993). Plants respond to chemical elicitor treatments by activating a wide variety of protective mechanisms designed to prevent pathogen replication and spreading (Malolepsza and Rózalaska, 2005). The defense mechanisms include the fast production of reactive oxygen species (De Gara et al., 2003); alterations in the cell wall constitution; accumulation of antimicrobial secondary metabolites known as phytoalexins (Agrios, 2005); activation and/or synthesis of defense peptides and proteins (Castro and Fontes, 2005). In various plant species, resistance can be induced with elicitors such as SA and H<sub>2</sub>O<sub>2</sub> against a wide range of pathogens (Abdel-Monaim 2010 and Abdel- Monaim et al., 2012).

In the present study, it was planning to investigate the possibility of minimizing the infection with damping-off, root-rot and wilt diseases of fababean using biocontrol agents (T. viride and B. megaterium) and chemical inducers (SA,  $H_2O_2$ ) individually and/or in combinations as resistance inducer.

The obtained data in vitro revealed that both biocontrol agents and chemical inducers individually and combination caused significant reduction growth of all tested pathogenic fungi. The combination between biocontrol agents and chemical inducers were more inhibited growth than used any of them individually especially in case of SA + T. viride and SA + B. megaterium .on the other hand, all treatments caused significant reduction to both damping-off and root rot/wilt diseases and increased the survival plants ether in pots or field experiments, compared with the control treatments. The combination between biocontrol agents and chemical inducers were more effective to reduce damping -off, root rot/wilt severity and increased fresh and weights of survival plants than used of them individually. SA + T. viride and SA + B. megaterium were recorded the best results in this respect. These results are in agreement with those reported by several rehearses (Metwally, 2004 and Abdel-Monaim, 2010). Saikia *et al.* (2003) tested the efficiency of *P. fluorescence* with or without SA amendment in chickpea against Fusarium wilt infection. The application of *P. fluorescence* (pf4-92) with SA recorded highest protection of chickpea seedlings against wilting. Rajkumar *et al.* (2008) found that pepper seeds treated with inducers (SA and chitin) alone showed a moderate degree of plant protection against *R. solani.* However, the reduction in disease was more pronounced when inducers were applied with fluorescent pseudomonades (SE21 and RD41) Amendment with chitin alone enhanced biocontrol efficiency of both SE21 and RD41. However, amendment with SA alone or in conjunction with chitin showed a moderate effect on biocontrol efficiency of the antagonists.

Seed soaking application of biocontrol agents and chemical inducers individually or combination in both season s showed a significant increase in faba bean growth parameters, yield components and protein content in seeds. The combination between biocontrol agents and chemical inducers were recorded highly increased in all growth and yield parameters more than in used of them individually, especially SA+T. viride and SA+B. megaterium. These increases may be attributed to biotic and abiotic elicitors effect on physiological processes in plant such as ion uptake, cell elongation, cell division, enzymatic activation and protein synthesis. In this concern, low SA and H<sub>2</sub>O<sub>2</sub> doses enhanced growth and yield components of tomato (Abdel-Monaim et al., 2012), alfalfa (Morsy et al., 2011). Also, Bacillus megaterium and T. viride increased plant growth and yield components in many plant species such as chickpea (Abdel-Monaim, 2010), dry bean (Yobo et al., 2011) and alfalfa (Morsy et al., 2011). Also, these results show that the biocontrol efficiency of antagonists T. viride and B. megaterium may be stimulated by SA and H<sub>2</sub>O<sub>2</sub> resulting in a significant increase in their population density and antagonistic effect against tested pathogens.

In conclusion, soaking faba bean seeds in biocontrol agent and chemical inducers individually or in combination significantly reduced damping-off, root rot and increased survival plants either under green house or field conditions. Also, these treatments increased plant growth, yield components and protein content in seeds in field during both growing seasons (2010-2011 and 2011-2012). The combination between biocontrol agents and chemical inducers were better than used of

them individually especially SA+*T. viride* and SA+ *B. megaterium*.

#### REFERENCES

- Abdel-Kader, M.M., S. El-Mougy, Nehal and S.M. Lashin. 2011. Essential oils and *Trichoderma harzianum* as an integrated control measure against faba bean root rot pathogens. J. of Plant Protection Res. 51 (3):306-313.
- Abdel-Monaim, M.F. 2010. Integrated management of damping- off, root and/or stem rot diseases of chickpea with sowing date, host resistance and bioagents. Egypt. J. Phytopathol. 38 (1-2): 45-61.
- Abdel-Monaim, M.F., M.A. Abdel-Gaid and H.A.H. Armanious. 2012. Effect of chemical inducers on root rot and wilt diseases, yield and quality of tomato. International J. of Agric. Sci. (7): 211-220.
- Abdou, El-S., H.M. Abd-Alla and A.A. Galal. 2001. Survey of sesame root/rot/wilt disease in Minia and their possible control by ascorbic and salicylic acids. Assuit J. of Agric. Sci. 32(3):135-152.
- Agrios, G.N. 2005. *Plant Pathology*. 5th Ed. Academic Press, San Diego, USA.
- Barnett, HL. and B.B. Hunter. 1986. Illustrated Genera of Imperfect Fungi. 4 th Ed., Macmillan Publishing Co., New York.
- Booth, C. 1985. The genus *Fusarium*. Kew, Surrey Commonwealth Mycological Institute, 2<sup>nd</sup> Ed., 237 pp.
- Chaven j.K., L.S. Kute and S.S. Kadam. 1989. In: CRC Hand Book of World Legumes, Eds., SaLUNKHE, d.d., s.s.Kadam. Boca Raton, Florida, USA, CRC Press: 223-245.
- De Boer, W., S. Gerards, P.J.A. Klein, R. Gunnewiek and Modderman. 1999. Response of the chitinolytic microbial community to chitin amendments of dune soils. Biology and Fertility of Soils. 29: 170–177.
- De Gara, L., M.C. De Pinto and F. Tommasi. 2003. The antioxidant systems vis à vis reactive oxygen species during plant pathogen interaction. Plant Physiol. And Biochem. 41: 863-870.
- Dhingra, O.D. and J.B. Sinclair. 1985. Basic plant pathology methods. CRC, Boca Raton, Florida, USA.
- Gomez, K. A. and A. A. Gomez. 1984. Statistical procedures for Agricultural research. A. Wiley-Interscience Publication. New York. pp. 678.
- Guzzo, S.D., E.A. Back, E.M. Martins and W.B. Moraes. 1993. Crude expolysaccarides (EPS) from

- Xanthomonas campestris pv. maniohtis, Xanthomonas campestris pv. campestris and commercial xanthan gum as inducers of protection in coffee plants against Hemileiava statrix. J. Phytopathol. 139: 119-128.
- Harrier, L.A. and C.A. Watson. 2004. The potential role of ArbuscularMycorrhizal (AM) fungi in the bioprotection of plants against soil-borne pathogens in organic and/or other sustainable farming systems. Pest Manag. Sci. 60 (2): 149-157
- Jackson, M.L. 1973. Soil Chemical Analysis. Prentice Hall, New Delhi, India.
- Jarvis, W.R. 1988. Fusarium crown root rot of tomatoes. Phytoprotection. 69: 49-64.
- Liu, L., J.W. Kloepper and S. Tuzun. 1995. Introduction of systemic resistance in cucumber against Fusarium wilt by plant growth-promoting rhizobacteria. Phytopathology. 85:695-698.
- Malolepsza, U. and S.Rózalaska. 2005. Nitric oxide and hydrogen peroxide in tomato resistance. Nitric oxide modulates hydrogen peroxide level in O. hydroxyethylorutin-induced resistance to *Botrytis cinerea* in tomato. *Plant Physiol. Biochem.* 43: 623-635.
- Metwally, M.M.M. 2004. Resistance induction against diseases of faba bean crop. Ph.D. Thesis, Fac. Agric., Suez Canal Univ.
- Morsy, K.M.M. 2005. Induced resistance against damping-off, root rot and wilt diseases of lentil. Egypt. J. Phytopathol. 33 (23): 53-63.
- Morsy, K.M., M.F. Abdel-Monaim and M.M. Mazen. 2011. Use of abiotic and biotic inducers in controlling fungal diseases and improving growth in alfalfa. Australian J. of Basic and Applied Sci. 5(9): 816-824.
- MSTAT-C. 1991. A Software Program for the Design, Management and Analysis of Agronomic Research Experiments. Michigan State University. pp. 400.
- Castro, M.S. and W. Fontes. 2005. Plant defense and antimicrobial peptides. Prot. Peptide Lett. 12: 11-
- Muthomi, J.W., P. E.Oteino, G.N. Chemining Wa, J. H. Nderitu and J.M. Wagacha, 2007. Effect of legume root rot pathogens and fungicide seed treatment on nodulation and biomass accumulation. J. of Biological Sciences. 7 (7): 1163-1170.
- Rajkumar M., K.J. Lee and H. Freitas. 2008. Effects of chitin and salicylic acid on biological control

- activity of *Pseudomonas* spp. against damping off of pepper. South African J. of Botany. 74 (2008) 268–273.
- Roby, D., A. Gadelle and A. Toppan. 1987. Chitin oligosaccharides as elicitors of chitinase activity in melon plants. Biochemical and Biophysical Res. Communications. 143: 885–892.
- Saikia, R., T. Singh, R. Kumar, J. Srivastava, A.K. Srivastava, K. Singh and D.K. Arora. 2003. Role of salicylic acid in systemic resistance induced by *Pseudomonas fluorescens* against *Fusarium oxysporum* f. sp. *ciceri* in chickpea. Microbiological Res. 158:203–213.
- Saleh N. M., A. K. Hussan, L.J. Sabir and A.A. Aish. 2009.

- Evaluation the efficacy of baking yeast, some nutrients and salicylic acid to control *Macrophomina phaseolina* The Iraqi J. of Agric. Sci. 40 (6):9-16
- Sturz, A.V. and R. Christie. 2003. Beneficial microbial allelopathies in the root zone: the management of soil quality and plant disease with rhizobacteria. Soil and Tillage Res. 72 (2): 107-123.
- Yobo K.S., M.D. Laing and C.H. Hunter. 2011. Effects of single and combined inoculations of selected *Trichoderma* and *Bacillus* isolates on growth of dry bean and biological control of *Rhizoctonia solani* damping-off. African J. of Biotechnology. 10(44): 8746-8756.